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Chunbo Yuan^a; Daqing Zhao^b; Bing Zhao^c; Jiazuan Ni^b

^a Department of Chemical Engineering & Chemistry, Southeast University, National Laboratory of Molecular and Biomolecular Electronics, Nanjing, P. R. China ^b Laboratory of Rare Earth Chemistry & Physics, Changchun Institute of Applied Chemistry, Academia Sinica, Changchun, P. R. China ^c Key Laboratory of Molecular Spectra and Structure, Jilin University, Changchun, P. R. China

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NMR AND FT-RAMAN STUDIES ON THE INTERACTION OF
LANTHANIDE IONS WITH SPHINGOMYELIN BILAYERS

Key Words: NMR, FT-Raman, Sphingomyelin Bilayers, Lanthanide ions

Chunbo Yuan¹, Daqing Zhao², Bing Zhao^{3*} and Jiazuan Ni²

¹National Laboratory of Molecular and Biomolecular Electronics,
Department of Chemical Engineering & Chemistry, Southeast University,
Nanjing 210096, P.R.China

²Laboratory of Rare Earth Chemistry & Physics, Changchun Institute of
Applied Chemistry, Academia Sinica, Changchun 130022, P.R.China

³Key Laboratory of Molecular Spectra and Structure, Jilin University,
Changchun 130023, P.R.China

ABSTRACT:

The interactions of lanthanide ions with sphingomyelin bilayers have been studied by using 2D NOESY spectroscopy and FT-Raman spectroscopy methods. The results indicate that lanthanide ions, as well as divalent calcium, combine mainly to the phosphate group in the polar headgroup and do not change the conformation of O-C-C-N⁺ backbone in the choline group of sphingomyelin bilayers. The polar headgroup is still

* To whom correspondence should be addressed.

extending parallel to the bilayer surface and O-C-C-N⁺ group is still in its gauche conformer.

INTRODUCTION

With the extensively application of rare earth in agriculture, medicament, especially the application of Gd-DTPA as nuclear magnetic resonance image reagent in clinical practice(1), the studies on the toxicology and the interactive mechanism of rare earth ions and their complexes in biological body, as well as the study on the use as informative probes in stead of divalent calcium ion in biological and biochemical research have attracted intensive concern(2).

Phospholipids bilayers have served as a model of biomembrane in research in last two decades. The effects of metal ions on the polar headgroup of bilayers are the focus(3-6) because the polar head-groups are located in the intermediary position between the hydrophobic and hydrophilic regions in the membrane bilayers in aqueous solution and therefore, the conformational changes would cause the structural changes of lipids bilayers and thus significantly influence the physiological function of membrane protein. Controversial conclusions were drawn from different experiments(3-5). We have used 2D NOESY and Raman spectroscopies in the investigation of the effects of trivalent lanthanide ions and their complexes on the conformation of polar headgroup of dipalmitoylphosphatidylcholine (DPPC) bilayers(7). To extend this work, we report here the results of 2D NOESY and FT-Raman spectroscopies studies on the influences of lanthanide ions on sphingomyelin (SPM, a major component of several biological tissues such as brain and nerve cells) bilayers which have identical polar headgroup as DPPC bilayers.

EXPERIMENTAL:

Sphingomyelin was commercially obtained from Sigma Chemical Co.. D_2O (99.8%), $CaCl_2$ (A.R.) were products of Beijing Chem. Plant. Ln_2O_3 were products of YaoLong Chemical Plant. Deuteriochloride(DCl) was obtained from Aldrich Chemical Co. Inc. All other reagents were reagent grade.

$LnCl_3$ solutions were prepared by dissolving their oxides into DCl solution for NMR experiments or in HCl solution for Raman measurements and eliminating the excess acid. pH of these solutions were around 5.4. The concentrations were determined by EDTA titrations, using arsenazo as indicator. Sphingomyelin liposomes were prepared as described elsewhere(8). The samples for Raman measurements were prepared as described in Ref.(9).

NMR experiments were carried out on Varian Unity 400 Spectrometer at 50 °C. T_1 values were measured by using the inversion recovery sequence. 2D NOESY spectra were performed with the pulse sequence $(\pi/2-t_1-\pi/2-\tau_m-\pi/2-t_2)n$. To suppress the J cross peaks, 10% random was introduced in mixed time(10).

FT-Raman spectra were recorded on Bruker RFS-100 FT-Raman spectrometer equipped with a InSb detector. Excitation was provided by a 1.064 μm CW Nd:YAG laser. The power of laser beam was 100 mW. Experimental temperature was maintained at 18 °C.

RESULTS AND DISCUSSION:

1. The Coordination Site of Lanthanide Ions to Sphingomyelin Bilayers

Table 1 shows the T_1 values of sphingomyelin bilayers with the presence and absence of Gd^{3+} . Gd^{3+} is a paramagnetic relaxation reagent.

TABLE 1. The spin-lattice times(T_1) and assignments of ^1H in sphingomyelin bilayer in absence and presence of Gd^{3+} .

Chemical Shift ppm	Assignment	$T_1(\text{sec})^*$	
		0.1 M SPM	0.01 M SPM + 0.02 mM Gd^{3+}
0.88	CH_3	0.85	0.83
1.28	$(\text{CH}_2)_n$	0.71	0.67
2.00	CH_2CON	0.70	0.65
3.22	$\text{N}(\text{CH}_3)_3$	0.43	0.32
3.66	CH_2N	0.42	0.33
4.29	CH_2OP	0.36	00.1
5.29	$\text{CH}=\text{CH}$	0.70	0.60

*The average of three times measurements, $\text{SD} = \pm 0.03 \text{ s}$

The scalar contribution of this ion to spin-lattice relaxation time(T_1) is negligible. Hence, the spin-lattice relaxation rate is proportional to r^{-6} , where r is the internuclear distance between the metal ion and nuclear in question. The greatest decrease in spin-lattice relaxation time was found in Table 1 of proton atoms near the phosphate moiety in polar headgroup. Therefore, we concluded that Gadolinium(III) ion was combined to the phosphate group of the polar headgroup of sphingomyelin bilayers. The lanthanide induced shifts(LIS) of some other lanthanide ions to the various protons of sphingomyelin molecule were also measured as shown in Table 2. The largest LIS data were observed at the protons close to phosphate group. These data were in accordance with T_1 values. Therefore, lanthanide ions combine mainly to the phosphate group in the polar headgroup of sphingomyelin bilayers.

TABLE 2 The lanthanide induced ^1H chemical shift(ppm) of sphingomyelin bilayers*

	Ce^{3+}	Pr^{3+}	Nd^{3+}	Eu^{3+}
$\text{N}(\text{CH}_3)_3$	0.06	0.16	0.18	- 0.11
CH_2N	0.10	0.25	0.21	- 0.24
$\text{CH}_2\text{OP}(\text{choline})$	0.20	0.46	0.30	- 0.57
CH_2OP	0.16	0.21	0.22	- 0.40
CHOCO	0.01	0.05	0.09	- 0.08

* 10 mmol.dm $^{-3}$ Ln^{3+} / 20 mmol.dm $^{-3}$ SPM

2. The Effect of Metal Ions on the Conformation of O-C-C-N $^+$ backbone

As described above, lanthanide ions were combining to the polar headgroup region of bilayers, concisely, to the phosphate group. The effect of lanthanide ions on the conformation of polar head-group is of great interest. We are involved in using 2D NOESY and FT-Raman spectroscopies studies on this issue.

Fig.1 shows the 2D NOESY spectra of sphingomyelin bilayers with or without the presence of lanthanide ion Eu^{3+} . In Fig.1A, sphingomyelin bilayers with the absence of Eu^{3+} ion, we found that proton of $\text{N}(\text{CH}_3)_3$ in polar headgroup has cross-peak not only with the methylene groups of choline group but also with the methylenes of hydrocarbon chains. As we known, the cross relaxation occurred through two pathways: spin diffusion and close spatial interaction. Experimental performed by Xu et al(11) with deuterated lipids had shown that cross relaxation is not mediated by spin diffusion, but by close spatial interaction. This means that the spatial distance between the protons of $\text{N}(\text{CH}_3)_3$ and methylene groups of hydrocarbon chains is within 5 Å ($\tau_m = 500$ ms) which allows the

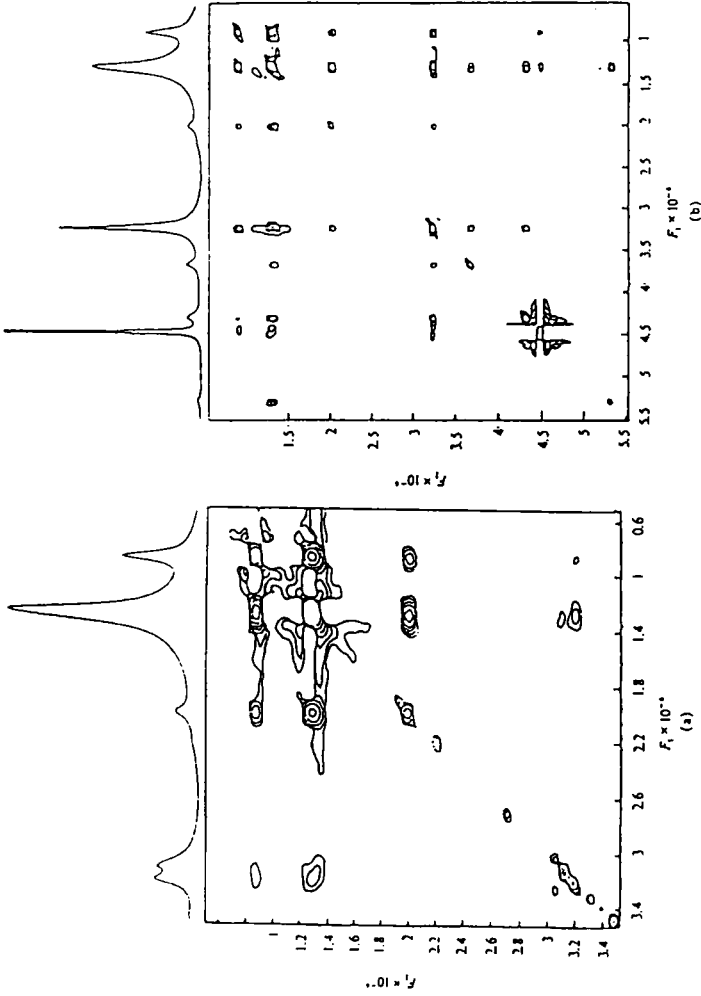


Fig. 1. 2D proton NOESY spectra of sonicated sphingomyelin liposomes (A) and sphingomyelin liposomes in the presence of Eu^{3+} ion (B). The molar ratio of Eu^{3+} /SPM is 1:5. The mixing time, τ_m is 500 ms and data set consisted of 512 blocks with 512 data points per block. Each block represents 32 acquisition.

development of NOE cross-peak between the protons in two groups. This is in agreement with that the polar headgroup of sphingomyelin bilayers in liquid-crystalline state is extending parallel to the membrane surface. In the presence of Eu^{3+} ions, a paramagnetic shift ion, which can only combine to the outer surface of sphingomyelin bilayers, and thereby only shifting the proton signal of outer surface to high field, two peaks of $\text{N}(\text{CH}_3)_3$ were observed in Fig. 1 and both of them have cross-peaks with the methylene groups of hydrocarbon chains. This strongly demonstrates that the presence of lanthanide ions does not alter the spatial distance between $\text{N}(\text{CH}_3)_3$ and methylene groups of hydrocarbon chains. Therefore, the polar headgroup in the presence of lanthanide ions is still extending parallel to the membrane surface.

The Raman spectra of sphingomyelin bilayers in 600-950 cm^{-1} regions with or without the presence of lanthanide ions and divalent calcium ion were shown in Fig.2. The characteristic bands observed in this region are 717, 763, 770, 875 and 889 cm^{-1} . The Raman band at 717 cm^{-1} has been assigned to C_4N^+ totally symmetric stretching mode(12). The band at 763 cm^{-1} is the result of OPO diester symmetric stretch(13), while 875 and 889 cm^{-1} have been assigned to carbon-carbon stretch in fluid-like and crystallinity state of interfacial region(13). The predominant changes of spectral feature in Fig.2 are the shift of 763 cm^{-1} to lower wavenumbers and 770 cm^{-1} to higher. Two bands occurred of OPO diester symmetric stretch may be resulted from different length of hydrocarbon chains in sphingomyelin molecules. When at higher concentration of lanthanide ions, the lower band at 763 cm^{-1} disappeared. This result demonstrates that the binding site of metal ions to bilayers is the phosphate moiety of polar

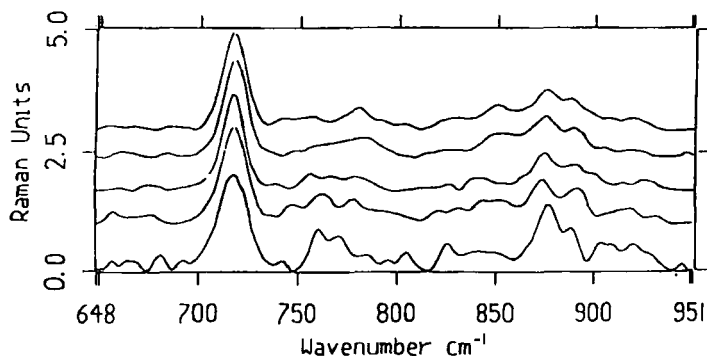


Fig.2 Raman spectra of sphingomyelin bilayers with or without the presence of metal ions in 650-950 cm^{-1} region. The lipid was dispersed in : a. Distilled water, b. 10 mM LaCl_3 solution, c. 40 mM LaCl_3 solution, d. 0.1 M LaCl_3 solution, e. 0.2 M CaCl_2 solution.

headgroup. This is in accord with the results of ^1H -NMR spin-lattice time and lanthanide induced chemical shift measurements. The band at 717 cm^{-1} that originated from the totally symmetric C-N stretching mode of C_4N^+ group, characteristic of O-C-C- N^+ backbone in its gauche conformation(12) remains invariant in the presence of La^{3+} and Ca^{2+} ions. No new band observed at 770 cm^{-1} that originated from the totally symmetric C-N stretching mode of C_4N^+ group when the backbone of O-C-C- N^+ in its trans conformation(12). The constancy of both the intensity and the frequency of 717 cm^{-1} band and no 770 cm^{-1} band occurred indicate that no conformational change occurred in polar headgroup of sphingomyelin bilayers combined with rare earth ions and calcium ion. The conformation of choline group is still in its gauche form which allows the polar headgroup extend parallel to the surface of sphingomyelin bilayers. Another apparent change observed in this region is the increase of the intensity ratios of

I_{889}/I_{875} in the presence of metal ions. The peak intensity ratio of I_{889}/I_{875} was used to monitor the physical state changes in interfacial region(13). The increase of the intensity ratio of I_{889}/I_{875} suggested that the interfacial region of sphingomyelin bilayers be in an ordered state. Certainly, This conclusion may be dependent on the further investigation.

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REFERENCES:

1. Lauffer R.B. Chem. Rev., 1987, 87:901.
2. Evans C.H. Biochemistry of Lanthanide, Plenum Press, New York, 1990.
3. Gally H.U., Nievesberg W., and Seelig J. Biochemistry, 1975, 14:3647.
4. Brown M.F., Seelig J. Nature, 1977, 269:721.
5. Hauser H., Phillips M.C., Levine B.A., Williams R.J.P. Nature, 1976, 261:390.
6. Akutsu H., Nagamori T. Biochemistry, 1991, 30:4510.
7. Yuan C.B., Zhao D.Q., Zhao B., Wu Y.J., Liu J.Z., Ni J.Z. Langmuir, Accepted
8. Bystrov V.F., Dubrovina N.I., Barsulkev L.I., Bergelson L.D., Chem. Phys. Lipids, 1971, 3:343.
9. Akutsu H., Suezaki Y., Yoshikawa W., Kyogoku Y., Biochim. Biophys. Acta, 1986, 854:213.
10. Zhao D.Q., Gao X.F., Pei F.K., Ni J.Z. Chinese Sci. Bull., 1993, 38:1094.
11. Xu Z.C., Cafico D.S., Biophys. J., 1986, 49:779.
12. Akutsu H., Biochemistry, 1981, 20:7359.
13. Brown E.B., Brown K.G., Ladjadj M., J. Phys. Chem., 1987, 91:3436.

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